

MELAMPOLIDES FROM *MAGNOLIA GRANDIFLORA*

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Abstract—Two isomeric melampolides, melampomagnolide A and melampomagnolide B, were isolated from the newly formed leaves of *Magnolia grandiflora*. Their stereochemical structures were established by spectroscopic means, and by chemical correlation with soulanganolide A and parthenolide, respectively.

INTRODUCTION

In a continuation [1] of our study of the C-14 partially oxidized melampolides of family Magnoliaceae, examination of the newly formed leaves of *Magnolia grandiflora* L yielded two novel compounds melampomagnolide A (1) and melampomagnolide B (2). This paper describes the isolation, structure elucidation and correlation of 1 and 2 with soulanganolide A (3) and parthenolide (4), respectively.

RESULTS AND DISCUSSION

An ethanolic extract of the newly formed leaves of *M. grandiflora* yielded the hitherto unreported melampolides melampomagnolide A (1) and the more polar melampomagnolide B (2), upon solvent partitioning then repeated chromatography. Melampomagnolide A (1), $C_{15}H_{20}O_4$, mp 177–178°, showed IR absorption bands and 1H NMR signals similar to those reported [1] for soulanganolide A (3), except for the absence of the characteristic C-1 olefinic proton. The presence of a C-1(10) epoxide group was suggested by the ^{13}C NMR signals at δ 62.6 (d) and 63.3 (s). Also, unlike soulanganolide A (3) [1], the hydroxymethyl group appeared in the 1H NMR spectrum as a pair of doublets ($J = 6.0$ and 12.0 Hz) centred at δ 3.50 and 3.95. This pattern was attributed to geminal coupling with additional coupling with the hydroxyl proton, as it was sharpened up to a pair of doublets ($J = 12.0$ Hz) upon deuterium exchange.

The stereochemical structure of melampomagnolide A (1) was confirmed by monoepoxidation of soulanganolide A (3) using *m*-chloroperbenzoic acid, the product was indistinguishable from melampomagnolide A (1). Since epoxidation would be expected to proceed peripherally [2], the stereochemistry at both C-1 and C-10 should be R.

Melampomagnolide B (2), $C_{15}H_{22}O_4$, mp 174–175°, could only be separated from melampomagnolide A (1) by flash chromatography [3]. Its 1H NMR, ^{13}C NMR signals (Table 1), and IR absorption bands were similar to

those reported for parthenolide (4), except for the absence of the C-14 methyl group, and the presence instead, of a hydroxymethyl group (see Experimental). In addition, the endocyclic proton (H-1) appeared in the 1H NMR spectrum as a broad triplet at δ 5.75 ($J = 7.0$ Hz). Its deshielded position relative to that of parthenolide (4) suggested a *cis* relationship with the hydroxymethyl group [1].

The structure of melampomagnolide B (2) was further confirmed by acetylation to yield the acetate 5 and manganese dioxide oxidation to the α,β -unsaturated aldehyde 6 with 1H NMR and ^{13}C NMR signals (Table 1) in agreement with their proposed structures. The stereochemical structure of 2 was unambiguously established by the regio- and stereo-specific allylic oxidation [4] of parthenolide (4) with selenium dioxide and *t*-butyl hydroperoxide. Melampomagnolide B (2) was the major product of this reaction (79%) with only a trace isolated of the aldehyde 6.

EXPERIMENTAL

Mps uncorr, IR 7–9% soln in $CHCl_3$ unless otherwise specified, 1H NMR 60 or 90 MHz, $CDCl_3$, TMS as int standard, ^{13}C NMR 15.03 MHz, $CDCl_3$, TMS as int standard. The plant material was collected in July 1980 on the campus of The University of Mississippi, University, Mississippi, U.S.A., and was identified by Professor Thomas Bullen, Biology Department of the same university. Parthenolide (4) and soulanganolide A (3) used in this investigation were isolated as previously reported [5, 1, respectively].

Isolation of melampomagnolides A (1) and B (2) The dried and powdered newly formed tender leaves (1.5 kg) of *M. grandiflora* were extracted by cold percolation with 95% EtOH. The extract (102 g) was partitioned between H_2O and $CHCl_3$, and the $CHCl_3$ solubles were further partitioned between *n*-hexane and 10% aq MeOH. Chromatography of the methanolic fraction (29 g) on silica gel using $CHCl_3$ as solvent gave a crystalline fraction (176 mg) that was flash chromatographed on silica gel using MeOH– C_6H_6 (1:9) (R_f values 0.20 and 0.27 on TLC) as solvent to give the following two compounds in the order of their elution.

Melampomagnolide A (1), crystallized from Et_2O to give fine needles (49 mg), mp 177–178°, $[\alpha]_D -19^\circ$ ($CHCl_3$, c 0.12), IR $\nu_{max}^{CHCl_3}$ cm^{-1} 3620 and 3490 (OH), 1758 (CO) and 1662 and 1670 (C=C), 1H NMR ($CDCl_3$) δ 1.85 (3H, s, Me-4), 2.52 (1H, s, exch OH), 3.50 and 3.95 (2H, each dd, $J = 6.0$ and 12.0 Hz,

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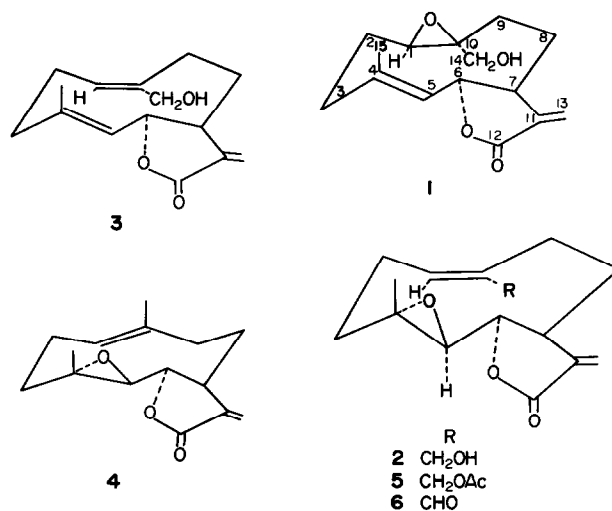


Table 1 ^{13}C NMR data for compounds 1 and 2 and their derivatives (15 03 MHz, CDCl_3 , TMS as int standard)

C	1	2	5	6	
1	62.6 (<i>d</i>)	126.8 (<i>d</i>)	130.6 (<i>d</i>)	153.5 (<i>d</i>)	
2	27.3 (<i>t</i>)*	24.2 (<i>t</i>)*	25.7 (<i>t</i>)*	22.5 (<i>t</i>)*	
3	24.2 (<i>t</i>)*	36.9 (<i>t</i>)*	36.6 (<i>t</i>)*	36.1 (<i>t</i>)*	
4	142.4 (<i>s</i>)	60.3 (<i>s</i>)	59.9 (<i>s</i>)	59.5 (<i>s</i>)	
5	124.4 (<i>d</i>)	63.3 (<i>d</i>)	63.3 (<i>d</i>)	63.1 (<i>d</i>)	
6	79.5 (<i>d</i>)	81.4 (<i>d</i>)	81.1 (<i>d</i>)	81.3 (<i>d</i>)	
7	46.4 (<i>d</i>)	42.8 (<i>d</i>)	42.6 (<i>d</i>)	42.3 (<i>d</i>)	
8	27.3 (<i>t</i>)*	25.7 (<i>t</i>)†	24.5 (<i>t</i>)†	25.0 (<i>t</i>)†	double intensity
9	34.6 (<i>t</i>)	23.7 (<i>t</i>)†	23.8 (<i>t</i>)†	25.0 (<i>t</i>)†	
10	63.3 (<i>s</i>)	139.8 (<i>s</i>)‡	130.0 (<i>s</i>)	144.0 (<i>s</i>)	
11	139.9 (<i>s</i>)	139.0 (<i>s</i>)‡	139.0 (<i>s</i>)	138.4 (<i>s</i>)	
12	170.1 (<i>s</i>)	169.8 (<i>s</i>)	169.4 (<i>s</i>)‡	169.3 (<i>s</i>)	
13	119.1 (<i>t</i>)	120.2 (<i>t</i>)	120.0 (<i>t</i>)	120.6 (<i>t</i>)	
14	64.8 (<i>t</i>)	65.4 (<i>t</i>)	66.8 (<i>t</i>)	195.0 (<i>d</i>)	
15	17.4 (<i>q</i>)	18.0 (<i>q</i>)	17.9 (<i>q</i>)	17.9 (<i>q</i>)	
Acetate C=O	—	—	170.6 (<i>s</i>)‡	—	
Acetate Me	—	—	20.9 (<i>q</i>)	—	

Assignments are based on comparison with reported chemical shift values for related compounds. See refs [1, 6, 7]

*, †, ‡ Assignments bearing the same superscript in the same column may be interchangeable

CH_2OH), 4.60 (1H, *t*, $J = 10.0$ Hz, H-6), 5.25 (1H, *d*, $J = 10.0$ Hz, H-5), 5.45 (1H, *d*, $J = 3.0$ Hz, H-13) and 6.17 (1H, *d*, $J = 3.0$ Hz, H-13), ^{13}C NMR (CDCl_3) Table 1, MS m/z 264 [$\text{M}]^+$ (< 1%) (Found C, 68.25, H, 7.77 $\text{C}_{15}\text{H}_{20}\text{O}_4$ (264) requires C, 68.16, H, 7.63%)

Melampomagnolide B (2), crystallized from Et_2O to give prisms (39 mg), mp 174–175°, $[\alpha]_{\text{D}} -46^\circ$ (CHCl_3 , c 0.18), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3475 (OH), 1748 (CO) and 1671 (C=C), ^1H NMR (CDCl_3): δ 1.50 (3H, *s*, Me-4), 2.80 (1H, *d*, $J = 9.5$ Hz, H-5), 3.84 (1H, *t*, $J = 9.5$ Hz, H-6), 4.12 (2H, *s*, CH_2OH), 5.62 (1H, *d*, $J = 3.0$ Hz, H-13), 5.70 (1H, *t*, $J = 9.0$, H-1) and 6.28 (1H, *d*, $J = 3.0$ Hz, H-13), ^{13}C NMR (CDCl_3) Table 1, MS m/z 264 [$\text{M}]^+$ (1%). (Found C, 69.35, H, 7.72 $\text{C}_{15}\text{H}_{20}\text{O}_4$ (264) requires C, 68.16, H, 7.63%)

Both melampomagnolide A (1) and melampomagnolide B (2)

were found only in newly formed tender leaves, although they were not detected in all collections made. For other constituents isolated and previously reported see ref [6]

***m*-Chloroperbenzoic acid oxidation of soulangianolide A (3) to melampomagnolide A (1)** Compound 3 (100 mg) was stirred in CHCl_3 soln for 4 hr with 79 mg of *m*-chloroperbenzoic acid (peroxy content 80%). The mixture was worked up in the usual manner [5] to give 93 mg of 1 as fine needles from Et_2O , identical with the natural material (mp, mmp, IR and ^1H NMR)

Oxidation of melampomagnolide B (2) to the aldehyde (6) Compound 2 (50 mg) was stirred in CH_2Cl_2 soln with 500 mg of MnO_2 overnight. Filtration and evaporation of the filtrate provided the aldehyde 6 as a colourless oil homogeneous on TLC (R_f 0.75 on silica gel with $\text{EtOH}-\text{CHCl}_3$, 1/19), $[\alpha]_{\text{D}} -73^\circ$ (CHCl_3 , c 0.41); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} 1678 (CHO) and no OH bands,

$^1\text{H NMR}$ (CDCl_3) δ 1.57 (3H, s, Me-4), 2.75 (1H, d, $J = 9.5$ Hz, H-5), 3.81 (1H, t, $J = 9.5$, C-6), 5.55 (1H, d, $J = 3.0$ Hz, H-13), 6.20 (1H, d, $J = 3.0$ Hz, H-13), 6.70 (1H, t, $J = 9.0$ Hz, H-1) and 9.52 (1H, s, CHO), $^{13}\text{C NMR}$ (CDCl_3) Table 1, MS m/z 262 $[\text{M}]^+$ (< 1%) (Found C, 68.57, H, 7.02 $\text{C}_{15}\text{H}_{18}\text{O}_4$ (262) requires C, 68.68, H, 6.92)

Acetylation of melampomagnolide B (2) to 5 Compound 2 (60 mg) was stirred with 2 ml $\text{C}_5\text{H}_5\text{N}-\text{Ac}_2\text{O}$ (1:1) for 24 hr. Usual work-up provided 5 (69 mg) as colourless prisms from isopropyl ether, mp 143–144°, $[\alpha]_D^{25} -27^\circ$ (CHCl_3 , c 0.12), IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$ same pattern as for 2 with a band at 1727 (AcO), $^1\text{H NMR}$ (CDCl_3) same pattern as for 2 with a signal at δ 2.05 (3H, s, AcO) and an AB system centred at δ 4.52 (2H, dd, $J = 12.5$ Hz, $-\text{CH}_2\text{OAc}$), $^{13}\text{C NMR}$ (CDCl_3) Table 1, MS m/z 306 $[\text{M}]^+$ (< 1%) (Found C, 66.69, H, 7.27 $\text{C}_{17}\text{H}_{22}\text{O}_5$ (306) requires C, 66.65, H, 7.24%)

Oxidation of parthenolide (4) to melampomagnolide B (2) Compound 4 (248 mg), *tert*-BuOOH (0.50 ml of 70% soln in H_2O), SeO_2 (56 mg) were stirred in 10 ml of CH_2Cl_2 for 8 hr. Flash chromatography [3] of the product on silica gel using EtOH– CHCl_3 (1:19) yielded 206 mg of 2 identical with the

natural product (mp, mmp, IR and $^1\text{H NMR}$), 7 mg of a product identical with 6 and 17 mg of unreacted 4

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ESSENTIAL OILS OF SOME AMAZONIAN MIKANIA SPECIES

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Abstract—Terpenoid constituents were identified in the essential oils of *Mikania banisteriae* and *M. congesta*. *M. amara* contains aliphatic alcohols, aldehydes and acids besides *p*-cymene and thymol

INTRODUCTION

Mikania banisteriae DC (called salsa branca) and *M. congesta* DC grow wild in Peixe-Boi and Bujaru, State of Pará, while *M. amara* Willd, known as 'cipó catinga', is distributed throughout Brazil and is esteemed for a number of medicinal properties, including use against fever, whooping cough and rheumatism [1]. Flavonoids [2, 3], sesquiterpenoid lactones [4, 5], kaurenoid diterpenes [6] and triterpenoids [7, 8] have been reported from several species of *Mikania*. Mono- and sesquiterpenes were identified in the essential oil of *M. mucrantha* [9]. As part of an ongoing study of the essential oils of Amazonian plants, we have analysed the volatile components of *M. amara*, *M. banisteriae* and *M. congesta* by GC/MS

RESULTS AND DISCUSSION

The essential oil of *M. amara* exhibited a chemical composition quite different from that of *M. banisteriae* and *M. congesta*. In these two latter species only mono- and sesquiterpenoids were found. The monoterpene α -pinene (43.3%) is the major component in the oil of *M. banisteriae* while limonene, β -cubebene, β -caryophyllene, germacrene B and an unidentified oxygenated sesquiterpene comprise 70% of the oil of *M. congesta*. The essential oil from *M. amara* is dominated by the presence of dodecanal, 1-dodecanol and tetradecanal, in addition to a large number of other aliphatic alcohols, aldehydes and acids.

Identification of most of the components was accomplished by comparison of both the mass spectrum and